

## Bio-inputs: An Alternative to Achieve Sustainable Agriculture Bioinsumos: Una Alternativa para Lograr una Agricultura Sostenible

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### SUMMARY

Agriculture involves the cultivation and domestication of plants, but different agricultural practices worldwide have led to the intensive use of natural resources, synthetic fertilizers and pesticides, as well as the use of heavy machinery and irrigation technologies. This has had negative consequences, mainly through the reduction of biodiversity, the appearance of resistant pests, imbalances in agroecosystems and harmful environmental effects. In view of this, research has focused on sustainable agriculture, whose production is economically, socially and environmentally acceptable. To achieve this, the application of bio-inputs formulated with biotechnological techniques from microorganisms, plants, compounds and extracts thereof, capable of improving crop yield and health, has been proposed. Therefore, the objective of this chapter is to show the advances in the production of bio-inputs, their characteristics and application results since they can gradually replace the use of synthetic inputs, reducing the pollutants generated by this sector, in addition to taking advantage of waste from various sectors to incorporate them into the circular economy. It is necessary to continue with research to integrate potential species in the formulation of bioproducts and to deepen in metaomics and bioinformatics with impact and improvement of agricultural activities.

**Index words:** *biofertilizers, biopesticides, biostimulants, biostabilizers, microbial inoculants.*

### RESUMEN

La agricultura involucra el cultivo y domesticación de plantas, pero las diferentes prácticas agrícolas a nivel mundial han ocasionado el uso intensivo de recursos naturales, de fertilizantes y pesticidas sintéticos, así como el empleo de maquinarias pesadas y tecnologías de riego. Provocando consecuencias negativas, principalmente con la disminución de la biodiversidad, aparición de plagas resistentes, desequilibrios en los agroecosistemas y efectos perjudiciales en el medio ambiente. Ante esto, se han realizado investigaciones enfocadas hacia una agricultura sostenible, cuya producción sea económica, social y ambientalmente aceptable. Para lograrlo, se ha propuesto la aplicación de bioinsumos formulados con técnicas biotecnológicas a partir de microorganismos, plantas, compuestos y extractos de estos; capaces de mejorar el rendimiento y sanidad de los cultivos. Por lo anterior, el objetivo del presente capítulo es mostrar los avances en la producción de bioinsumos, sus características y resultado de aplicación; ya que pueden sustituir paulatinamente el uso de insumos sintéticos reduciendo los contaminantes que genera este sector, además de aprovechar residuos de diversos sectores para incorporarlos a la economía circular. Es necesario continuar



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con la realización de investigaciones para integrar especies potenciales en la formulación de bioproductos y profundizar en las metaómicas y la bioinformática con impacto y mejoramiento de las actividades agrícolas.

**Palabras clave:** *biofertilizantes, bioplaguicidas, bioestimulantes, bioestabilizadores, inoculantes microbianos.*

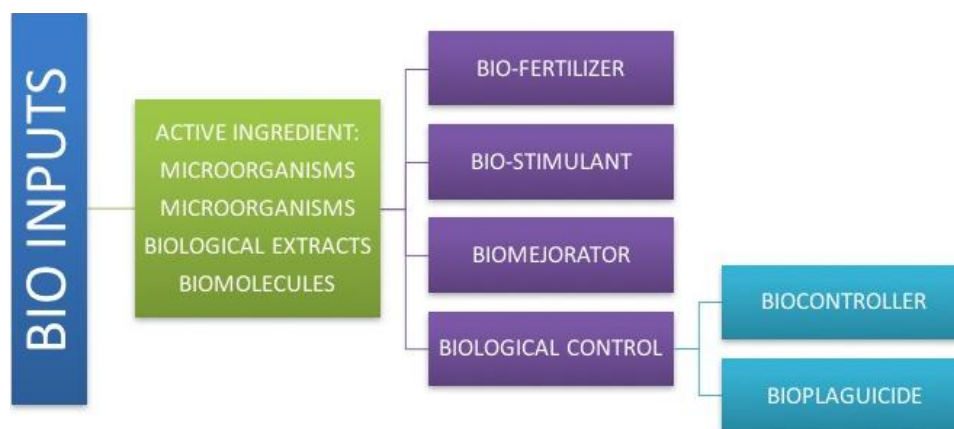
## INTRODUCTION

Agriculture is a millenary activity that has been perfected over time to treat the soil and cultivate the land with the main objective of producing food and satisfying human needs. Due to the great demand for agricultural products worldwide, natural resources have been exploited, but poor cultivation practices and the massive use of chemical compounds have caused soil erosion and poor-quality products that affect the health of consumers, as well as a decrease in biodiversity, the appearance of resistant pests, imbalances in agroecosystems and harmful effects on the environment (Grageda-Cabrera, Díaz, Peña, and Vera, 2012). In view of this, research has been conducted focused on economically and socially acceptable agricultural production. To achieve this, the production and application of bio-inputs have been proposed (Mamani and Filippone, 2018).

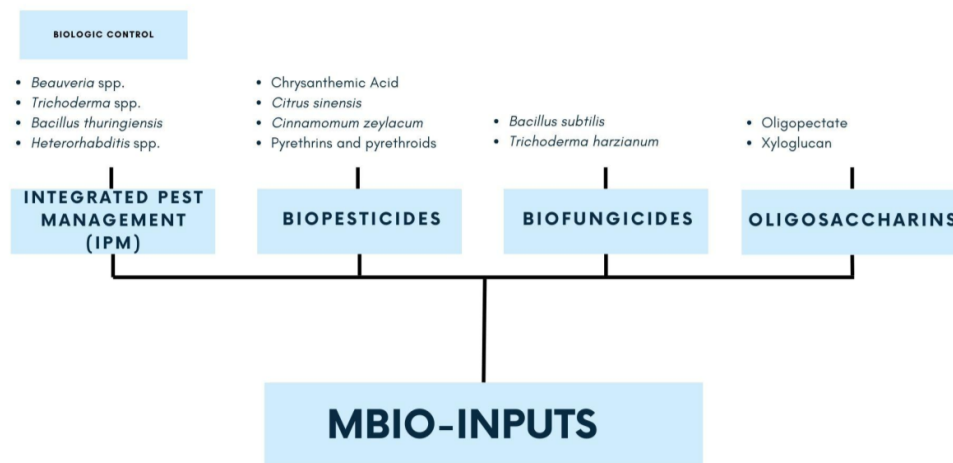
These are biological products formulated from compounds and extracts of microorganisms or plants, or live microorganisms, capable of improving productivity (or yield), quality, and health of soils and crops, resulting in environmentally friendly Figure 1 (Fillipone *et al.*, 2017).

Biotechnology is used for their formulation since they use strategies that arise from the study and characterization of the different interactions of plants with their environment; they can be of plant or microbial origin. Depending on their effect on the plant, they are classified as biofertilizers (biostimulants, microbial inoculants, biostabilizers, organic fertilizers, humus, and guano) and biopesticides (microbiocides, bioinducers of plant defense against pests and diseases, and biorepellents) (Figure 2). In addition to bioinputs, biotechnology has contributed to the *in vitro* cultivation of plants in laboratories and greenhouses and the commercial exploitation of transgenic plants in different agroecological zones of the world (Christeson and Sims, 2011; Logan and Rabaey, 2012). Molecular engineering in the agricultural area has allowed obtaining plants resistant to insect pests and pathogens; improved products, such as fruits with prolonged storage periods with better nutritional properties (higher content of proteins, oils, amino acids) and industrial improvements (higher content of fruit solids).

The increase in population density has created a challenge for agriculture because it is necessary to supply the growing demand for food products; food security is related to climate change, the conditions of which have worsened, with alterations expected to become increasingly drastic. To address this problem, it is necessary to increase the implementation of sustainable agriculture and generate alternatives or products that allow mitigation of the pollution produced in agriculture using residues and the reduction of the use of chemical fertilizers and pesticides. In addition, it is necessary to generate new plants that are better adapted and more productive (Barajas, 2017; Chaves, 2021).



**Figure 1.** Types of bio-inputs used in agriculture.



**Figure 2. Integration of the main bio-inputs and active ingredients.**

Therefore, the purpose of this review is to show the advances in the production of bioinputs, their characteristics and the results of their application, and thus expand our knowledge of these alternative biotechnological products for agriculture.

## DEVELOPMENT

### Technological Innovations with Application in Agriculture

Currently, there are several products on the market derived from biotechnological research that present some activity or improvement in plant development and that are emerging as an alternative in agricultural activities. The use of microorganisms, such as fungi, bacteria, viruses; or their metabolites and insects, to eliminate or reduce the damage caused by harmful organisms that affect crops have been proposed as biological control agents in Integrated Pest Management (IPM), their purpose is to reduce chemical agents, having biofungicides and bioinsecticides as an ecological option (Viera-Arroyo *et al.*, 2020).

Also, carbohydrates have been used, among which are oligosaccharins, Oligogalacturonides, and Xyloglucans, which induce a defensive response and resistance in plants, in addition to stimulating plant growth and development (Falcón-Rodríguez, Costales, González, and Nápoles, 2015; Lara, Costales, and Falcón, 2018).

On the other hand, the use of chitosan stimulates plant root growth, acts as an antimicrobial agent to combat pathogens in the soil, presents an activity to control the deterioration of post-harvest products during storage and the export process; as well as in the elaboration of fertilizer coatings for controlled release (Katiyar, Hemantaranjan, Singh, and Bhanu, 2014; Bauer, Villegas, and Zucchetti, 2022).

Nowadays, co-products, by-products, and solid and liquid organic residues (raw material) from the fishing, agricultural, and industrial sectors (food, sugar, tequila) contain bioactive compounds and minerals of agricultural interest. These residues are used in bioprocesses that employ methods such as aerobic biodegradation, fermentation, hydrolysis, and extraction (macroalgae). The products obtained can be considered as organic fertilizer and biostimulant (depending on their composition of macronutrients, micronutrients, amino acids, microorganisms, and bioactive compounds), whose effect ranges from the improvement of growth and development in plants to the control of biotic and abiotic stress in crops so that they can grow without difficulties (Florez-Jalixto, Roldán, Omote-Sibina, and Molleda, 2021). Each of these bioproducts is detailed below.

### Use of Biological Control in Integrated Pest Management (IPM)

Biological control uses living organisms or their metabolites to eliminate or reduce damage caused by harmful organisms. Biological control agents (BCA), such as fungi, bacteria, viruses, and insects, are being developed worldwide to reduce the population of pests affecting crops (Table 1). The use of entomopathogenic fungi has gained importance in recent years because they do not cause damage to the environment and health (Viera-Arroyo *et al.*, 2020).

**Table 1. Main organisms used in biological control with application to crops of agricultural importance.v**

Microorganism	Application, action, or effect in agriculture	Reference
<i>Beauveria</i> spp. <i>B. bassiana</i> <i>B. brongniartii</i> .	Management of numerous veterinary and forestry agricultural arthropod pests. It is implemented in one or more flooding applications of large quantities of aerial conidia in dry or liquid formulations. Mass production is mainly practiced by solid-state fermentation to produce hydrophobic aerial conidia, which are the main active ingredient of mycoinsecticides. Infection of the fungus begins with the adhesion of the conidia to the cuticle of the host; when these germinate, they attach to the cuticle, forming a hyphae that allows it to penetrate the insect with the help of hydrolytic enzymes (proteases, lipases, chitinases), mechanical pressure and other factors. The hyphal bodies produce blasto-spores that disperse throughout the host body, destroying tissues, evading the immune system, and producing toxins that contribute to death. It is used in a variety of crops for the control of whiteflies ( <i>Bemisia tabaci</i> ), mites ( <i>Tetranychus urticae</i> ), beetles ( <i>Gonipterus scutellatus</i> ), coffee berry borer ( <i>Hypothenemus hampei</i> ), banana borer ( <i>Cosmopolites sordidus</i> ), defoliating worm ( <i>Dione juno</i> ), thrips, aphids, bugs, leafhoppers.	(Wraight and Ramos, 2002; Humber, 2008; Lacey, Wraight, and Kirk, 2008; Malpartida-Zevallos, Narrea, and Dale, 2013; Ortiz-Urquiza and Keyhani 2013; Jaramillo, Montoya, Benavides, and Góngora, 2015; Mascarin and Jaronski, 2016; Espinel et al., 2018)
<i>Trichoderma</i> spp. <i>T. harzianum</i> . <i>T. asperellum</i> . <i>T. viride</i> . <i>T. atroviride</i> . <i>T. gamsii</i> . <i>T. hamatum</i> . <i>T. koningii</i> . <i>T. lignorum</i> . <i>T. polysporum</i>	The species of this fungus act as biofungicide, biofertilizer, growth promoter, and inducer of natural resistance; its application can be foliar before planting (in seeds or propagation material), incorporated into the soil during planting or transplanting (by means of irrigation). It is the most widely used biological fungicide in agriculture. The fungi live freely in the soil and root ecosystems, possessing antagonistic properties and therefore exerting biocontrol of phytopathogenic fungi such as <i>Fusarium</i> , <i>Rhizoctonia</i> , <i>Pythium</i> , <i>Sclerotium</i> , and <i>Phytophthora</i> , which affect crops of commercial interest such as rice, corn, onion, tomato, bean, wheat, cocoa, and quinoa. Indirectly, it competes for space and nutrients, producing antibiotics or stimulating plant growth and defense mechanisms; directly, it can control mycoparasitosis through the production of different metabolites.	(Woo et al., 2014; Charoenrak and Chamswarn, 2016; Tirado-Gallego, Lopera, and Ríos 2016; Leon-Ttacca, Ortiz, Condori, and Chura, 2018)
<i>Bacillus thuringiensis</i>	Entomopathogenic bacteria must be ingested and act by releasing toxins or penetrating midgut cells before invasion of the hemocele to multiply in the insect carcass. After causing death, the new progeny of the bacteria leave to infect new hosts. Gram-positive bacteria such as <i>Bacillus thuringiensis</i> (Bt) and insect pathogens of the genera <i>Paenibacillus</i> and <i>Lysinibacillus</i> have the ability to form spores. In contrast, Gram-negative bacteria do not form spores and include isolates of the genera <i>Serratia</i> , <i>Yersinia</i> , <i>Photobacterium</i> , <i>Chromobacterium</i> . Several entomopathogenic bacteria can be produced by fermentation. <i>B. thuringiensis</i> active strains act on Lepidoptera, Coleoptera and Diptera. Due to its greater stability in the production, formulation, and field application processes, using Cry spores and protein crystals as the active ingredient, it is the species most widely used on a large scale for the control of insect pests. Species such as <i>B. subtilis</i> , <i>B. pumilus</i> , <i>B. licheniformis</i> , and <i>B. amyloliquefaciens</i> have been successfully implemented in commercial formulations developed mainly for the control of fungal diseases in grasses, oilseeds, fruit trees, vegetables, and ornamentals.	(Grijalba, Hurst, Ibarra, Jurat, and Jackson, 2018; Villareal-Delgado et al., 2018)
<i>Heterorhabditis</i> spp., <i>Steinernema</i> spp.	Entomopathogenic nematodes are soil-dwelling organisms that interact as important regulators of their ecosystem and are used as controllers of lepidopteran pests and other types of insect pests in commercially important crops. Their use is based on their ability to adapt to new environments and adverse conditions, resistance to chemicals, high specificity for insects, harmlessness to the environment and mammals, and compatibility with other entomopathogens. The species of the genus <i>Heterorhabditis</i> present a cephalic structure like a spike in the juvenile infectives, which gives them an additional way of entering the insect; this characteristic makes them more infective with respect to other species, since in addition to penetrating through the natural openings (anus, mouth, spiracles), they can also do so by perforating the cuticle.	(Castillo, Gallegos, Asaquibay, and Oña 2011; Gianfelici, Bertolotti, and Cagnolo, 2014; López-Llano and Soto-Giraldo, 2016; Rumbos and Athanassiou, 2017; Sánchez, Valle, Pérez, Neira, and Calderón, 2019)
Plant extracts <i>Jatropha curcas</i> , <i>Azadirachta indica</i> , <i>Melia azedarach</i> , <i>Citrus aurantifolia</i> , <i>Eucalyptus globulus</i> , <i>Piper nigrum</i> , <i>Punica granatum</i> , <i>Salix safsaf</i>	They are derived from some parts or active ingredients of plants; among these are attractants, hormones, plant and insect growth regulators, enzymes, and chemical signaling substances of importance in the plant-insect relationship. These pesticides contain phytochemicals or secondary metabolites such as terpenes, phenols (tannins and coumarins), alkaloids, cyanogenic glycosides, sulfur compounds (thiophenes) and flavonoids (rotenone); these substances cause repellency, inappetence, prevent oviposition, act as a barrier by their taste, inhibit growth, are toxic to nematodes, mites and insects, release cyanide, act as enzyme inhibitors.	(Jannet, H-skhiri, Mighri, Simmonds, and Blaney, 2001; Martínez, 2012; Nava-Pérez, García, Camacho, and Vázquez, 2012)
Aqueous seaweed extract <i>Padina gymnospora</i> (Kützing) Sonder, <i>Sargassum latifolium</i> (Turner) C. Agardh <i>Hydroclathrus clathratus</i> (C. Agardh) M. Howe., <i>Sargassum vulgare</i>	<i>In vivo</i> application of seaweed powder as a soil amendment decreases the percentage of root rot disease caused by <i>Fusarium solani</i> on <i>Solanum melongena</i> L. (eggplant). The antifungal activity of extracts of the algae <i>Laminaria digitata</i> , <i>Undaria pinnatifida</i> (Harvey) Suringar, and <i>Porphyra umbilicalis</i> Kützing against the postharvest pathogens <i>Botrytis cinerea</i> , <i>Monilinia laxa</i> , and <i>Penicillium digitatum</i> in strawberries, peaches and lemons is highlighted by inhibiting mycelial growth and conidial germination. The aqueous and methanolic extract of <i>Sargassum vulgare</i> has shown antifungal potential by inhibiting the mycelial growth of <i>Pythium aphanidermatum</i> , reducing the disease observed in potato tubers cv. spunta.	(De Corato, Salimbeni, De Pretis, Avella, and Patrino, 2017; Ibraheem, Hamed, Abd Elrhman, Farag, and Abdel, 2017; Ammar et al., 2017)

As a result of the application of biotechnology in agriculture, several products have been developed and placed on the market. The formulations are contemplated in an Integrated Pest Management (IPM) plan, such as biopesticides (insects, mites, nematodes) and biofungicides, or they are incorporated into crops as biofertilizers or biostimulants to promote plant development through efficient assimilation of nutrients.

Biopesticides (Table 2) are products that contain microbial control agents (entomopathogens), entomophagous organisms, organic compounds as the active ingredient, or metabolites of the microorganism that are extracted by procedures that do not alter their composition. These products have demonstrated their toxic activity for pest control, cause minimal environmental damage, and do not leave toxic residues in food, so they are not considered a risk factor in humans (Biokrone, 2023).

A biofungicide is a compound whose active material is an antagonistic microorganism, a beneficial microorganism such as fungi or bacteria that acts on pathogenic plant diseases. Biofungicides (Table 3) are an alternative to chemical fungicides (Biokrone, 2023).

### Use of Oligosaccharins

Oligosaccharins (Table 4) are natural polysaccharides and oligosaccharides that are part of the cell walls of plants and microorganisms such as fungi; the main sources of raw material for their large-scale preparation are agricultural by-products and the exoskeleton of crustaceans discarded from the fishing industry (Falcón-Rodríguez *et al.*, 2015). They can be endogenous or exogenous.

Chitosan is also considered as oligosaccharin, it is a natural polymer derived from chitin (the main component of the shell of crustaceans, it is also found in the cell wall of fungi and the exoskeleton of some insects, it is the second most abundant natural biopolymer); It acts as a plant root growth enhancer, an antimicrobial agent to combat pathogens in the soil, a gel that controls the deterioration of post-harvest products during storage and during the export process, as well as in the elaboration of fertilizer coatings for controlled release (Katiyar *et al.*, 2014; Bauer *et al.*, 2022).

### Development of Biofertilizers and Stimulants

The indiscriminate use of chemical fertilizers is one of the main problems in agriculture, leading to the loss of soil fertility and contamination of water bodies. As an alternative, biofertilizers are a biotechnological tool that consists of applying microorganisms that help to improve the availability of nutrients in the soil and thus reduce the effects caused by the excess of fertilizers. Their use in different crops has evidenced positive effects on soil fertility; they are applied to seeds, plant surfaces, or soil to colonize the rhizosphere or the interior of the plant and promote growth by increasing the supply or availability of primary nutrients to the host plant.

Plant growth-promoting rhizobacteria (rhizosphere-dwelling bacteria) are phytostimulants containing bacterial auxins capable of inducing root elongation. Several genera such as *Acetobacter*, *Acidithiobacillus*, *Aminobacter*, *Arthrobacter*, *Azoarcus*, *Azospirillum*, *Azotobacter*, *Azolla*, *Bacillus*, *Burkholderia*, *Clostridium*, *Enterobacter*, *Gluconoacetobacter*, *Pseudomonas*, *Serratia* and *Sphingomonas*; have demonstrated their growth promoting capacity, so they are among the main genera used for the formulation of commercial biofertilizers, which are applied in cereal and vegetable crops. Their main function is to mobilize nutrient availability based on their biological activity, help recover lost microbiota and generally improve soil health (Malusá, Sas-Paszt, and Ciesielska, 2012; Ismail, Mohamed, Khattab, and Sherif, 2014; Velasco-Jiménez, Castellanos, Acevedo, Aarland, and Rodríguez, 2020).

The impact of chitosan, mycorrhizal fungi, and humic acids on the growth of bell pepper (*Capsicum annuum* L.) varieties under protected conditions has been reported, finding that the three biostimulants increased seed germination between 11.66 and 16.67%, while emergence was enhanced by humus and chitosan. Humic acids produced taller plants and larger diameter stems. When humic acids were applied, higher yields were achieved due to increased length, diameter, weight, and fruits per plant (Reyes-Pérez *et al.*, 2021). On the other hand, the agronomic response of *Vigna unguiculata* L. Walp (beans) was greater when seeds were inoculated with a strain of the arbuscular mycorrhizal fungus *Rhizophagus irregularis* (Tamayo, Riera, Terry, Juárez, and Rodríguez, 2019).

A similar case has been observed in evaluating the effect of different nutritional management on yield and internal and external quality of tomato fruits. The combination of 50% mineral fertilizer - bioproducts (partial organic nutrition treatment), allowed obtaining an adequate tomato yield without affecting the internal and external quality of the harvested fruits. Therefore, the bioproducts contributed to partially replacing the mineral fertilization required by the tomato crop (Terry-Alfonso, Ruiz, and Carrillo, 2018).



**Table 2. Biopesticides marketed for pest control.**

Biotechnology product	Active ingredient/ biological	Action	Manufacturer or distributor	Reference
Cridor®	Chrysanthemic acid and pyritic acid	<i>Chrysanthemum cinerariaefolium</i> extract is effective for the control of pests such as <i>Frankliniella occidentalis</i> , <i>Lygus</i> sp. <i>Pentatrichopus fragaefolii</i> , <i>Scirtotrips perseae</i> , <i>Idona minuenda</i> , <i>Brevicoryne brassicae</i> , <i>Plutella xylostella</i> , <i>Bemisia tabaci</i> , <i>Bactericera cockerelli</i> in vegetables, strawberries and avocado.	Syngenta Agro, S.A. de C.V.	(Syngenta, 2023)
Candor®	<i>Citrus sinensis</i> <i>Cinnamomum zeylacum</i>	Botanical acaricide and insecticide based on citrus and cinnamon extracts that acts on <i>Tetranychus urticae</i> , <i>Bemisia tabaci</i> , <i>Brevicoryne brassicae</i> ; used for IPM on vegetables, cucurbits, and strawberries.	Syngenta Agro, S.A. de C.V.	(Syngenta, 2023)
Pirecris®	Pyrethrins and pyrethroids	An incapacitating effect known as knock down that acts by contact and ingestion, producing an immediate paralysis of the insect and the death of the pest (decreases the population). It is indicated for the control of pests such as aphids and whiteflies; in avocado, strawberries, vegetables, cucurbits, broccoli, cabbage, cauliflower, rape, kiwi, and grapes.	SEIPASA MEXICO, S.A. DE C.V.	(Seipasa, 2023)
Cinrerate™	Cinnamon extract oil	It acts by contact, penetrating and destroying the soft parts of the body. Insects (avocado thrips) and mites are exposed to loss of body fluids, causing dehydration and death. It is applied on vegetables, cucurbits, strawberries, citrus and avocado.	SEIPASA MEXICO, S.A. DE C.V.	(Seipasa, 2023)
BIO BT®	<i>Bacillus thuringiensis</i> var. <i>kurstaki</i> and <i>Bacillus thuringiensis</i> var. <i>aizawai</i>	During the sporulation, it produces delta endotoxins ( $\delta$ -endotoxins), components of the protein crystals formed, which once activated in the intestine of the insect by enzymatic effect under conditions of alkaline pH of the stomach of the larvae. These cause osmotic imbalances that break the intestinal wall of <i>Helicoverpa zea</i> , causing death due to lack of food or septicemia.	AGRHUSA Agrobiológicos	(AGRHUSA, 2023)
AGRONEEM®	Neem seed extract oil (azadirachtin)	The components of neem extract, especially azadirachtin, enter the body of the insect ( <i>Bemisia tabaci</i> ) either by contact or ingestion, suppressing the activity of ecdysone, preventing the insect from molting or causing malformations that lead to death or sterility. In addition, it has a phagodisulative effect, discourages oviposition of females, inhibits chitin formation and acts as a repellent of larvae and adult insect pests.	AGRHUSA Agrobiológicos	(AGRHUSA, 2023)
Benemite®	Geraniol + Citronellol + Nerolidol + Farnesol	Broad spectrum acaricide contains pheromones that alter sexual and feeding behavior, interferes in the molting process of immature stages such as nymphs and protonymphs, blocks the spiracles causing asphyxia of the mite and decomposition of the chitin (rupture of the exoskeleton of the mite). It can be used on strawberries, vegetables, cucurbits, corn, avocado, and rose.	Arysta LifeScience México, S.A. de C.V.	Arysta LifeScience, 2023
EPA 90®	Soybean vegetable oil	The insecticidal action is based on its property of forming a layer that completely covers nymphs and adults, blocking the spiracles (gas exchange), which causes suffocation or asphyxiation of the insects. As a highly lipophilic product, it alters the chorion, cell membrane, and integument of eggs, nymphs, and adults, causing their death by desiccation. Recommended for <i>Trips tabaci</i> , <i>Bemisia tabaci</i> , <i>Diaphorina citri</i> , <i>Tetranychus urticae</i> , <i>Paraleyrodes perseae</i> , <i>Frankliniella occidentalis</i> ; in chives, citrus, strawberries, avocado, and vegetables.	Biokrone, S.A. de C.V.	Biokrone, 2023
NEMOVER	Extracts of fig ( <i>Ricinus communis</i> ), oregano ( <i>Origanum vulgare</i> ) and pine ( <i>Pinus pinaster</i> )	It acts by contact with a broad spectrum of control and immediate effect on the populations of soil pests and nematodes that attack different vegetable crops. By adhering to the substrate and roots, it impregnates the particles with its active ingredients, inhibiting the presence of eggs on the surface where the product had contact, contains the development of the nematode ( <i>Meloidogyne</i> spp.) in the second larval stage and causes intoxication by feeding on the epidermal tissues. Controlling nematodes and soil pests allows healthy and abundant roots to develop.	ECOPROTECTO, S.A. DE C.V.	Agroorganicos nacionales, 2023

**Table 3. Biofungicides marketed for the control of phytopathogens.**

Biotechnology product	Active ingredient/ biological	Action	Manufacturer or distributor	Reference
Cinnerate™	Cinnamon extract oil	It exerts a curative action, cinnamic aldehyde inhibits mycelial growth and prevents germination of fruiting bodies. It affects mitochondrial oxidative phosphorylation and induces profound changes in the enzymatic activity of the cell membrane of the pathogen (powdery mildew, downy mildew), causing dehydration and the disappearance of foliar symptoms. Crops: Avocado, vegetables, strawberries, citrus, cucurbits, tobacco, solanaceae, kiwifruit, grapes.	SEIPASA MÉXICO, S.A. DE C.V.	Seipasa, 2023
Fungisei®	<i>Bacillus subtilis</i> strain IAB/BS03	It acts by contact; the selected strain of <i>Bacillus subtilis</i> could produce lipopeptide antibiotics (iturins, surfactins, and phengicins), which prevent pathogenic spores from germinating, altering the growth of the germ tube of the spores and thus inhibiting the attachment of the pathogen (powdery mildew, gray mold) to the leaf; in bell pepper, grape, and blueberry.	SEIPASA MÉXICO, S.A. DE C.V.	Seipasa, 2023
Seipa System	Acacia Extract	Strengthens the plant's defensive system: Together with proteins, it generates self-defense molecules that inhibit the hydrolytic enzymes of phytopathogenic fungi from penetrating plant cells. Mechanical resistance is produced in the plant due to the association of silicon with cell wall components. It controls <i>Fusarium</i> and <i>Botrytis</i> in vegetables, grapes, and strawberries.	SEIPASA MÉXICO, S.A. DE C.V.	Seipasa, 2023
Baktillis®	<i>Bacillus subtilis</i> spores and metabolites	It acts as a physical barrier on the plant surface preventing adherence and germination of phytopathogens. The lipopeptides it contains perforate the mycelial membranes and fungal spores, preventing their development. In addition, the metabolites subtilin, bacitracin, and toximicin act directly on the spores and mycelium of phytopathogens. Recommended for the control of <i>Fusarium</i> , <i>Botrytis</i> , <i>Colletotrichum</i> , among others; present in strawberries, citrus, and vegetables	Biokrone, S.A. de C.V.	Biokrone, 2023
NatuControl®	Spores of <i>Trichoderma harzianum</i>	Directly attacks and parasitizes the structures of phytopathogenic fungi, including those inside the plant. Induces resistance that activates plant defense mechanisms. Produces substances and metabolites that act directly on spores or mycelium preventing the germination and development of phytopathogenic fungi. Rapidly colonizes the rhizosphere of the plant, preventing the germination, growth, and development of phytopathogenic fungi such as <i>Phytophthora</i> , <i>Rhizoctonia</i> , and <i>Fusarium</i> , among others. Recommended for: strawberries, vegetables, citrus, cucurbits.	Biokrone, S.A. de C.V.	Biokrone, 2023
Iodus 40® (Vacciplant)	Laminarin based product (extracted from seaweed)	Activates the intrinsic protection of the plant (strengthens the cell wall, phytoalexin production, and production of PR protein). Recommended for use in vegetables, cucurbits, and strawberries, among others, to control <i>Botrytis</i> , <i>Alternaria</i> , <i>Xanthomonas</i> , ash, mildew, and anthracnose.	Laboratorios GOEMAR SAS (Francia).	Arysta Life Science, 2023

The use of mycorrhizae has been related to crops that have a seedling stage, as occurs in horticulture; these fungi are important in nutrient uptake, protection under water stress conditions, and prevention against pathogens. The mycorrhizal association is relevant in soil stability and in reducing the negative effects of microbial competition, in the establishment, survival, and growth of plants in the field, as well as in stimulating crop yields (Plana-Llerena, González, and Soto, 2016).

The use of algal biostimulants in plant crops can generate numerous benefits with reported effects, including improved rooting, higher crop and fruit yields, increased photosynthetic activity, and resistance to fungi, bacteria, and viruses. With *Padina gymnospora*, the growth of eggplant (*Solanum melongena* L.) in *Fusarium solani*-infected soils is improved (Sharma, Fleming, Selby, Rao, and Martin, 2014; Ibraheem, Hamed, Abd Elrhman, Farag, and Abdel, 2017).

Biofortifiers and biostimulants (Table 5) are products based on microorganisms, organic matter, plant extracts, and oils, among others, whose application promotes and improves plant efficiency in the absorption and assimilation of nutrients and contributes to tolerance to biotic or abiotic stresses.

**Table 4. Products or research that has been carried out based on oligosaccharins with applications in the agricultural sector.**

Product	Application, action or effect in agriculture	Reference
Oligogalacturonides are located in the pectic portion of the cell wall of plants Oligopectate (pectin-derived oligosaccharide)	Increased color in the fruit of <i>Vitis vinifera</i> var. <i>Flame Seedless</i> (grape), related to increased anthocyanin content and possible induction of PAL (Phenylalanine ammonia lyase) mRNA expression.	(Ochoa-Villarreal, Vargas, Islas, González, and Martínez, 2011)
Xyloglucans: hemicelluloses that make up the primary cell wall structure of dicotyledonous and non-poaceous monocotyledonous plants. In dicotyledon seeds, they are part of the reserve polysaccharides	Induction of defensive response and resistance in plants, influencing plant growth and development. Tamarind seeds are rich in xyloglucans.	(Falcón-Rodríguez, Costales, González, and Nápoles, 2015; Lara, Costales, and Falcón, 2018)
Pectimorph®. Composed of biologically active carbohydrates. 55-61% galacturonic acid, which constitutes vegetable pectin, is present mainly in fruits.	Replaces growth regulators <i>in vitro</i> culture, promotes rooting in cuttings, stimulates crop growth and yield, and accelerates seed germination. Crops: Coffee, sugar cane, garlic, potato, cassava, tomato, banana, radish, carrot, lettuce, chard, spinach, soybean, bean, guava, citrus, grape, peach, fig, cinnamon, laurel, rose, orchid, lily, anthurium, bougainvillea, ficus, areca, thuja.	(Falcón-Rodríguez <i>et al.</i> , 2015; Lara <i>et al.</i> , 2018; Ramos <i>et al.</i> , 2013; INCA, 2023)
EcoMic®. Formulation based on Arbuscular Mycorrhizal Fungi, <i>Glomus cubense</i> strain	It increases the absorption capacity of soil nutrients and fertilizers, decreases the damage of some root and foliar pathogens, contributes to improving soil structure, and increases carbon sequestration in agroecosystems. Crops: Beans, corn, bananas, rice, soybeans, sorghum, vegetables, cotton, sunflower, citrus and fruit trees, grass species, flowers, and trees.	(Rivera <i>et al.</i> , 2020; INCA, 2023)
Quitomax®. Biostimulant based on natural biodegradable polymer, chitosan	It allows a reduction in the application of chemical pesticides on crops and increases from 10 to 30% the yield of plants; its combined application benefits the process of nitrogen fixation and growth in leguminous plants. Crops: corn, sorghum, potato, tomato, bell pepper, garlic, onion, soybean, bean and rice.	(Morales-Guevara, Dell Amico, Jerez, Hernández, and Martín, 2016; Rodríguez-Pedroso <i>et al.</i> , 2017; Gustavo-González <i>et al.</i> , 2021; INCA, 2023)
AzoFert®. Biofertilizer based on rhizobia ( <i>Rhizobium</i> ) and nodulation inducers	Direct nitrogen supply to the plant, promotes plant growth and crop yield, increases the utilization of soil nutrients, as well as the protection of the root system against certain diseases. It has been used in bean and soybean crops.	(Estrada <i>et al.</i> , 2017; Hernández and Salido 2019; Calero, Quintero, Pérez, Jiménez, and Castro 2020; INCA, 2023)
Coating of seeds with a 2% solution of chitosan	Positive effect on the microbiological development of the <i>Zea mays</i> (maize) plant, acting as a seed protector against pathogenic fungi under abiotic stress conditions (drought, humidity, acid pH, and alkaline pH)	(Lizárraga-Paulín, Torres, Moreno, and Miranda, 2011)
Crosslinking of chitosan in hydroxypropyl methylcellulose for the preparation of Neem oil coating for postharvest storage of pitaya ( <i>Stenocereus pruinosus</i> )	Application of chitosan-based antimicrobial coatings for postharvest preservation of pitaya (storage at 10 ± 2 °C, relative humidity of 80 ± 5%). This reduced weight loss and fungal contamination, increasing shelf life (up to 15 days).	(Hernández-Valencia, Román, Aguilar, Cira, and Shirai, 2019a)



**Table 5. Biofertilizers or biostimulants marketed to promote nutrient uptake and assimilation in plants.**

Biotechnology product	Active ingredient/biological	Action	Manufacturer or distributor	Reference
Isabion®	Organic matter (mixture of amino acids and peptides)	Fertilizers of natural origin are classified as an organic nutrient that supplies plants with amino acids and peptides to stimulate their vegetative development. Recommended for: vegetables, ornamentals, cucurbits, alliaceae and brassicas	Syngenta Agro, S.A. de C.V.	Syngenta, 2023
ALGACROP®	Seaweed 80%, Mixture of: <i>Ascophyllum nodosum</i> and <i>k</i> sp.	High content of amino acids, phytohormones, carbohydrates, and other organic compounds that promote plant growth and development; prevent abortion in the flowering stage; improve fruit quality and firmness; favor root development; and cover nutritional deficiencies by increasing the availability and absorption of elements. Crops: vegetables and tobacco.	AGRHUSA Agrobiológicos	(AGRHUSA, 2023)
ULTRAHUMI®	Humic acids, fulvic acid	Humic acids are the reverse of nutrients necessary for the plant, as these require organic matter as a carbon source. It helps the plant to avoid stress, stimulates bacterial growth in the rhizosphere, acts as a soil improver by increasing moisture retention, stimulates respiration, metabolism, and cation exchange capacity at the cellular level, reduces the harmful effects of toxic agents, favors the recovery of insoluble phosphorus and increases soil microbial life. Crops: vegetables and tobacco.	AGRHUSA Agrobiológicos	(AGRHUSA, 2023)
E-Microzime®	Primary microorganisms: <i>Azotobacter vinelandii</i> , <i>Clostridium pasteurianum</i>	Microbial enzyme complex contains a variety of soil microorganisms that help increase soil fertility and provide vital nutrients to all types of plants and crops.	MYDAGRO LLC.	(PLM® DEAO/ DIPO, 2023)
AmiKrone®	L-amino acids, peptides, and polypeptides of natural origin. Aspartic Acid, Glutamic Acid, Alanine, Amino Acids, Glycine, Lysine, Manganese, Organic Matter, Methionine, Proline, Zinc.	It is rapidly absorbed by plants either via foliar or root (depending on the form of application), acts as a biostimulant by triggering and enhancing numerous reactions of plant metabolism, provides vitamins and gives resistance in times of stress, also controls the water content inside, which gives it a prolonged life. It can be applied to vegetables, alliaceae, cucurbits, brassicas, legumes, grasses, fruit trees, ornamentals, and strawberries.	BIOKRONE, S.A. DE C.V.	Biokrone, 2023
AZSeed®	<i>Azospirillum brasilense</i> , <i>Azotobacter</i> spp., <i>Rhizobium</i> spp., <i>Bacillus</i> spp., Natural rooting agents (AIA) Inoculant and root bioactivator.	When applied to the seed and deposited on the soil or substrate, the bacteria contained in the product are activated and begin to reproduce rapidly, generating a synergism with the plant roots. The beneficial bacteria fix atmospheric nitrogen and give it to the plant. The metabolites (AIA) it contains ensure uniform seed germination and rapid rooting of the plants, thus generating vigorous plants. It can be used in grasses, vegetables, brassicas, alliaceae, fruit trees, citrus, legumes, oilseeds, strawberries, sugar cane, ornamentals, and agave.	BIOKRONE, S.A. DE C.V.	Biokrone, 2023
GLUMIX®	Spores of <i>Glomus geosporum</i> , <i>G. fasciculatum</i> , <i>G. constrictum</i> , <i>G. tortuosum</i> , <i>G. intraradices</i> , selected strains of vesicular arbuscular mycorrhizal fungi (VAM)	It invades the roots when they emit exudates or chemical compounds that stimulate the germination of the spores; inside the root, a series of dichotomous ramifications begin that give rise to an arbuscule, which occupies the cortical cells of the host. The hyphae and mycelium of the mycorrhizae penetrate the root of the plant, coming into contact with the ribosomes (root cells). Externally to the roots, the mycorrhizae emit large quantities of mycelium or hyphae that extend exploring a considerable volume of soil; in this way, the mycorrhizae contribute to the better development and growth of the plant. It is recommended for use in grasses, vegetables, brassicas, alliaceae, fruit trees, citrus, legumes, oilseeds, strawberries, sugar cane, ornamentals, and agave.	BIOKRONE, S.A. DE C.V.	Biokrone, 2023

## Main Mechanisms of Action of microorganisms used

In general terms, the mechanisms of action employed by some of the microorganisms used to produce biofertilizers are summarized in Figure 3.

## Genetic Improvement of Crops of Agricultural Interest

Another way in which biotechnology has contributed to the agricultural sector is the development of *in vitro* tissue culture methodologies, which allow the massive propagation of individuals of interest, making it possible to work with multiple individuals in a reduced space and controlled conditions, as well as the possibility of adding selection agents to the culture medium. Thus, it is also possible to obtain plants resistant to drought, herbicides, and diseases. Other important biotechnologies are medium and long-term conservation by cryopreservation (storage of plant tissues such as seeds, apices, buds, and cells under ultra-low temperature conditions, using liquid nitrogen [-196 °C]) (Chaves, 2021).

Micropropagation or *in vitro* propagation is one of the applications of plant tissue culture that contributes to accelerating the selection processes within genetic improvement programs. It has been applied for the genetic improvement of various crops. For example, sugarcane has been decisive in developing varieties resistant to biotic and abiotic factors and in increasing biomass and sucrose yields. In the *in vitro* regeneration of sugarcane plants, the cultivation of protoplasts, cells, callus, tissues, and various organs has been used (Castañeda-Castro *et al.*, 2014).

Soil salinity affects food production on a global scale by causing physiological and biochemical changes in plant metabolism that determine their survival and productivity under these conditions, for which plants have developed tolerance mechanisms. Genetic improvement offers an increase in the recovery of underutilized areas and in yields in areas where salinity is a limiting factor. The selection of salinity tolerant plants is a long process, so efficient indicators are sought for the early selection of genotypes with better agronomic performance (Lamz and Gonzalez, 2013).

Modern genetic improvement has two stages: the first is to obtain genetic variability and the second consists of selecting the genotypes thus obtained to develop varieties adapted to specific conditions. Crossing between different genotypes of the same species is the conventional tool for generating variability through classical breeding.

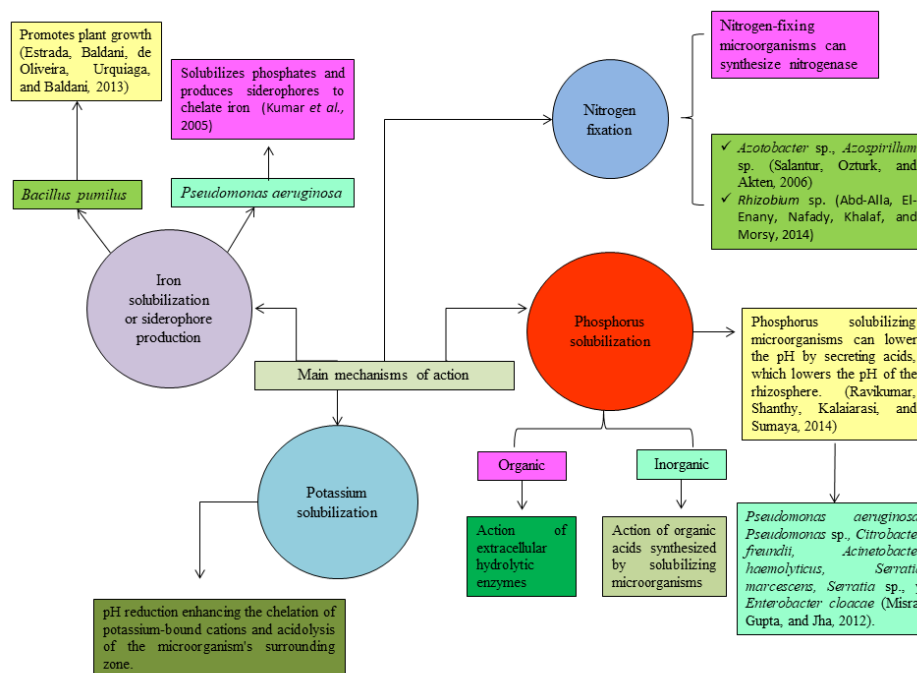


Figure 3. Summary of the main mechanisms of action of microorganisms used in agriculture.

Biotechnology makes it possible to make use of all existing genetic variability, circumvent the sexual barrier to incorporate genes from other species, integrate artificial genes, or edit the genome (making it possible to generate genetic variability). Another advantage is the possibility of introducing a single new gene in the genetically improved plant, preserving the rest of the genes of the original plant in its offspring. This technology has been used in crops to increase insect resistance and herbicide tolerance; organisms to which genetic material has been introduced in this way are called “genetically modified organisms” (GMOs).

In this sense, *Bacillus thuringiensis* (Bt) is a Gram-positive bacterium in *Bacillus* form, which produces Cry proteins with biological activity against insect pests. Due to these proteins, *B. thuringiensis* presents toxicity against insect-pest larvae of the orders Lepidoptera, Coleoptera, and Diptera, among others. It is common in the development of commercial products and transgenic plants based on Cry toxins in the agricultural sector, such as cotton and corn. The transformation of plants, through the incorporation of Cry protein-coding genes, confers protection to internal tissues, maintains a constant production of protein, and does not generate a harmful environmental impact (De Maagd, Bravo, and Crickmore, 2001).

Transgenic foods are those generated from genetic engineering and genome editing techniques. Golden Rice is a variety with nutritional improvement due to the increase in vitamin intake in which two genes are inserted through genetic engineering techniques, resulting in an increase in the amount of  $\beta$ -carotene in the grains. It is contemplated that this variety will serve to attenuate blindness in juvenile populations while it will reduce child malnutrition and malnutrition.

### Prospects for Achieving Sustainable Agriculture

Bioprospecting is the research conducted to identify species, varieties, genes, and valuable natural components of organisms in order to develop products for commercial purposes with current or potential use for their application in agriculture, industry, and the environment. Considering the diversity of microscopic organisms that inhabit the planet, as well as the multiple bioactive substances present in higher organisms; it is important to give continuity to bioprospecting, as this will allow the development of bioproducts, culture media or identify suitable substrates for the growth of beneficial micro and macro organisms for agriculture, such as *Trichoderma* (Páramo-Aguilera, Fonseca, Delgado, Cabistán, and Ríos, 2018), which is important for its ability to preserve and restore soil microbiota, for its potential for the synthesis and release of enzymes such as cellulases, xylanase, and chitinases, for the production of auxins and crop growth regulators, as well as a biological agent that can significantly reduce the incidence of diseases in crops under greenhouse conditions (Andrade-Hoyos *et al.*, 2023).

In this sense, modern agriculture is facing new challenges, integrating ecological and molecular approaches to achieve higher crop yields and minimize the impact on the environment. In order to generate higher yields, chemical pesticides are used, and the dose of synthetic fertilizers per unit area has increased significantly, inputs that can cause pollution, damage to health, and loss of soil fertility, becoming one of the most important concerns in agricultural production. To improve production without the use of fertilizers of synthetic origin and pesticides, research has been oriented towards the development of new biotechnologies: causing there to be a growing interest in beneficial soil microorganisms as these can promote plant growth and, in some cases, prevent infections of plant tissue by pathogens.

These challenges are determined by the demographic increase and the demand for raw materials for human and animal food, by preferences in consumption, by environmental and legal aspects, and by the globalization of the economy (Melgarejo, Romagosa, and Durán, 2014; Moreno, Carda, Reyes, Vásquez, and Cano, 2018).

Endophytic fungi are microorganisms that inhabit plant tissue without causing apparent symptoms of damage and are capable of producing bioactive metabolites that confer benefits to the host. With the purpose of obtaining secondary metabolites, it has been possible to isolate endophytic fungi from plants and soils, which have shown antimicrobial activities, as well as stimulating plant development. However, further research is still needed to provide information on the toxicity and safety of secondary metabolites so that they can be used as agrochemical agents. In addition, it is essential to conduct studies on the role that these metabolites may have in nature and the protection they provide to their host against pathogens and herbivores (Sánchez-Fernández *et al.*, 2013).

The use of basidiomycetes fungi in agriculture has aroused interest due to their ability to efficiently degrade aromatic and heterogeneous compounds because they present a nonspecific enzyme complex with oxidative activity against a wide variety of toxic and recalcitrant substances, and some species have shown antifungal, phytotoxic and nematicidal activity. In addition, basidiomycetes have the ability to efficiently degrade lignin (a process that involves the synergistic action of extracellular enzymes). They can also transform simpler substances

into environmental pollutants. For these reasons, they are promising species as biological controllers with an interest in agriculture; they can influence the control of plant diseases and thus become a potential biotechnological tool. The bioactivity of basidiomycetes has not been sufficiently investigated and constitutes a source for bioassays, suggesting new lines of development of different metabolites with a wide range of applications (Rojas, 2013).

On the other hand, macroalgae are a source of macro and microelements, bioactive compounds, growth regulators, and organic matter; they have nutritional value, stimulate plant defense against pathogens, improve soil characteristics, and increase agricultural production in a sustainable manner. Seaweed hulls are a resource with numerous possibilities for their use as raw material to produce flour, extracts, compost, and biotechnological products, with the aim of reducing the use of chemicals in soils. Therefore, it is transcendental to continue trials with algae species that have not been studied in order to know the compounds they contain and the mechanism of action in plants, in soils, and the activity on pathogens of agronomic importance, both in field and greenhouse conditions (Durán-Hernández, Uribe, Mateo, and González, 2022).

The production of organic fertilizers and biostimulants for the development of commercial formulas must first be developed at the laboratory scale, optimizing the working parameters to be scaled up, in addition to demonstrating the absence of phytotoxic substances and pathogenic microorganisms. In this sense, it is important to continue promoting sustainable strategies for more efficient and cost-effective waste disposal for the benefit of organic agriculture, aquaculture, and the environment, in addition to framing it in a circular economy model (Florez-Jalixto *et al.*, 2021).

On the other hand, research for breeding transgenic plants will continue to grow, and the long-term outlook is that transgenics will continue to be introduced into world agriculture, occupying larger areas and partially or totally replacing traditional varieties (Ardisana, Álvarez, Macías, Gainza, and 2016).

## CONCLUSION

Biotechnology techniques applied to agriculture are oriented towards sustainability since they focus on the development of products or innovations that reduce the pollutants generated by this productive sector. This may be due to the indiscriminate use of chemical fertilizers and insecticides or to the extensive generation of residues and leachates that accumulate on the surface, in the air, and in bodies of water. Thanks to biotechnology and microbiology, it has been possible to develop bio-inputs, the purpose of which is to gradually replace the use of synthetic products to reduce pest populations and control the spread of diseases that affect crops of agricultural interest. In addition to the use of waste from various sectors to incorporate them into what is known as the circular economy. However, it is still necessary to continue with research to incorporate potential species in the formulation of bioproducts, characterize them, and define their interaction with their environment to determine the degree of stimulation of reactions they cause in plants or other microorganisms, whether beneficial or pathogenic. It is also necessary to deepen research that contemplates meta-omics and bioinformatics with impact and improvement of agricultural activities.

## ETHICS STATEMENT

Not applicable.

## CONSENT FOR PUBLICATION

All authors have read and approved the final manuscript and have given their consent for publication.

## AVAILABILITY OF SUPPORTING DATA

Data sets used or analyzed during the current study are available from the corresponding author upon reasonable request.

## COMPETING INTEREST

The authors declare that they have no competing interests.

## FINANCING

Not applicable.

## AUTHORS' CONTRIBUTION

Conceptualization: J.A.C. Methodology: B.S.V.L., E.M.H.D., J.A.C., R.R.C. Research: R.S.C. Preparation of the original draft: R.S.C. Writing: revision and editing: J.A.C. and R.R.C.

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## REFERENCES

- Abd-Alla, M. H., El-Enany, A. W. E., Nafady, N. A., Khalaf, D. M., & Morsy, F. M. (2014). Synergistic interaction of *Rhizobium leguminosarum* bv. *viciae* and arbuscular mycorrhizal fungi as a plant growth promoting biofertilizers for faba bean (*Vicia faba* L.) in alkaline soil. *Microbiological research*, 169(1), 49-58. <https://doi.org/10.1016/j.micres.2013.07.007>
- AGRHUSA (2023). Agrobiológicos. Consultado el 28 de marzo, 2023, desde <http://www.agrhusa.com.mx/publico/principal/index.aspx?m=0>
- Agro orgánicos nacionales (2023). Productos de origen biológico. Consultado el 30 de marzo, 2023, desde <https://agrorganicosnacionales.com/53-categorias-productos/174-biologicos-menu>
- Ammar, N., Jabnoun-Khiareddine, H., Mejdoub-Trabelsi, B., Nefzi, A., Mahjoub, M. A., & Daami-Remadi, M. (2017). Pythium leak control in potato using aqueous and organic extracts from the brown alga *Sargassum vulgare* (C. Agardh, 1820). *Postharvest Biology and Technology*, 130, 81-93. <https://doi.org/10.1016/j.postharvbio.2017.04.010>
- Andrade-Hoyos, P., Rivera-Jiménez, M. N., Landero-Valenzuela, N., Silva-Rojas, H. V., Martínez-Salgado, S. J., & Romero-Arenas, O. (2023). Beneficios ecológicos y biológicos del hongo cosmopolita *Trichoderma* spp. en la agricultura: una perspectiva en el campo mexicano. *Revista Argentina de Microbiología*, 55(4), 366-377.
- Ardisana, E. F. H., Álvarez, S. P., Macías, R. M., & Gaínza, B. M. (2016). Perspectivas futuras e impacto social de las biotecnologías vegetales. *Alternativas*, 17(2), 44-51.
- Arysta LifeScience (2023). Productos. Consultado el 29 de marzo, 2023, desde <https://www.arystalifescience.com/eng-us/global-products.html>
- Barajas, L. N. A. (2017). Biofertilizantes: conceptos, beneficios y su aplicación en Colombia. *Ingeciencia*, 2(1), 65-76.
- Bauer, J. L., Villegas, L. F., & Zucchetti, A. (2022). Aplicaciones del quitosano en la agricultura, la industria y la salud: Applications of chitosan in agriculture, industry and health. *South Florida Journal of Environmental and Animal Science*, 2(2), 37-45. <https://doi.org/10.53499/sfjeasv2n2-001>
- Biokrone (2023). Biofortificante y bioestimulantes. Consultado el 27 de marzo, 2023, desde <https://biokrone.com/biofortificantes/>
- Calero-Hurtado, A., Quintero-Rodríguez, E., Pérez-Díaz, Y., Jiménez-Hernández, J., & Castro-Lizazo, I. (2020). Asociación entre AzoFert® y microorganismos eficientes como potenciadores del crecimiento y la productividad del frijol. *Revista de la Facultad de Agronomía*, 37(4), 387-409. [https://doi.org/10.47280/RevFacAgron\(LUZ\).v37.n4.04](https://doi.org/10.47280/RevFacAgron(LUZ).v37.n4.04)
- Castañeda-Castro, O., Gómez-Merino, F. C., Trejo-Téllez, L. I., Morales-Ramos, V., González-Arnao, M. T., Martínez-Ocampo, Y. M., ... & Pastelín-Solano, M. C. (2014). Aplicaciones del cultivo de tejidos vegetales en caña de azúcar. *Agro Productividad*, 7(2), 16-21.
- Castillo, C. C., Gallegos, P., Asaquibay, C., & Oña, M. (2011). *Guía de prospección y multiplicación de nematodos entomopatógenos para el control biológico de plagas en Ecuador*. Quito, Ecuador: INIAP.
- Charoenrak, P., & Chamswang, C. (2016). Efficacies of wettable pellet and fresh culture of *Trichoderma asperellum* biocontrol products in growth promoting and reducing dirty panicles of rice. *Agriculture and Natural Resources*, 50(4), 243-249. <https://doi.org/10.1016/j.anres.2016.04.001>
- Chaves, J. P. (2021). Biotecnología vegetal: mejoramiento de cultivos ante el cambio climático. *Investiga. TEC*, 14(42), 3-5.
- Christeson, L., & Sims, R. (2011). Production and harvesting of microalgae for wastewater treatment, biofuels, and bioproducts. *Biotechnology Advances*, 29(6), 686-702.
- De Corato, U., Salimbeni, R., De Pretis, A., Avella, N., & Patrino, G. (2017). Antifungal activity of crude extracts from brown and red seaweeds by a supercritical carbon dioxide technique against fruit postharvest fungal diseases. *Postharvest Biology and Technology*, 131, 16-30. <https://doi.org/10.1016/j.postharvbio.2017.04.011>
- De Maagd, R. A., Bravo, A., & Crickmore, N. (2001). How *Bacillus thuringiensis* has evolved specific toxins to colonize the insect world. *Trends in Genetics*, 17(4), 193-199. [https://doi.org/10.1016/S0168-9525\(01\)02237-5](https://doi.org/10.1016/S0168-9525(01)02237-5)
- Durán-Hernández, D., Uribe-Orozco, M. E., Mateo-Cid, L. E., & González-Mendoza, D. (2022). Potencial biotecnológico de las macroalgas en la agricultura. *Idesia*, 40(3), 81-88.
- Espinel, C. C., Torres, T. L. A., Villamizar, R. L. F., Bustillo, P. A. E., Zuluaga, M. M. V., & Cotes, P. A. M. (2018). Hongos entomopatógenos en el control biológico de insectos plaga. En P. A. M. Cotes (ed.) *Control biológico de fitopatógenos, insectos y ácaros: agentes de control biológico* (pp. 334-367). Colombia: Agrosavia. <https://doi.org/10.21930/agrosavia.investigacion.7402537>
- Estrada, G. A., Baldani, V. L. D., de Oliveira, D. M., Urquiaga, S., & Baldani, J. I. (2013). Selection of phosphate-solubilizing diazotrophic *Herbaspirillum* and *Burkholderia* strains and their effect on rice crop yield and nutrient uptake. *Plant and Soil*, 369, 115-129. <http://dx.doi.org/10.1007/s11104-012-1550-7>
- Estrada-Prado, W., Chávez-Suárez, L., Jerez-Mompie, E., Nápoles-García, M. C., Sosa-Rodríguez, A., Cordoví-Domínguez, C., & Celeiro-Rodríguez, F. (2017). Efecto del Azofert® en el rendimiento de variedades de frijol común (*Phaseolus vulgaris* L.) en condiciones de déficit hídrico. *Centro Agrícola*, 44(3), 36-42.



- Falcón-Rodríguez, A. B., Costales-Mené, D., González-Peña Fundora, D., & Nápoles-García, M. C. (2015). Nuevos productos naturales para la agricultura: las oligosacarinas. *Cultivos Tropicales*, 36, 111-129.
- Fillipone, M. P., Peto, D., de Los Ángeles, P., Grellet-Bournonville, C. F., Chalfoun, N. R., Tórtora, M. L., ... & Castagnaro, A. P. (2017). Bioproductos y sanidad vegetal. *Avance Agroindustrial*, 38, 24-32.
- Florez-Jalixto, M., Roldán-Acero, D., Omote-Sibina, J. R., & Molleda-Ordoñez, A. (2021). Biofertilizantes y bioestimulantes para uso agrícola y acuícola: Bioprocesos aplicados a subproductos orgánicos de la industria pesquera. *Scientia Agropecuaria*, 12(4), 635-651. <https://dx.doi.org/10.17268/sci.agropecu.2021.067>
- Gianfelicci, M., Bertolotti, M. A., & Cagnolo, S. R. (2014). Susceptibilidad de larvas de *Crocidosema aporema* (Walsingham, 1914) y *Anticarsia gemmatilis* Hübner, 1818, a tres aislados de nematodos entomopatógenos. *Revista de la Facultad de Ciencias Exactas, Físicas y Naturales*, 12, 71-76.
- Grageda-Cabrera, O. A., Díaz-Franco, A., Peña-Cabriales, J. J., & Vera-Núñez, J. A. (2012). Impacto de los biofertilizantes en la agricultura. *Revista Mexicana de Ciencias Agrícolas*, 3(6), 1261-1274.
- Grijalba, E., Hurst, M. Ibarra, J. E., Jurat-Fuentes, J. L., & Jackson, T. (2018). Bacterias entomopatógenas en el control biológico de insectos. En P. A. M. Cotes (Ed.). *Control biológico de fitopatógenos, insectos y ácaros: agentes de control biológico* (pp. 19296-333). Colombia: Agrosavia.
- Gustavo-González, L., Paz-Martínez, I., Boicet-Fabré, T., Jiménez-Arteaga, M. C., Falcón-Rodríguez, A., & Rivas-García, T. (2021). Efecto del tratamiento de semillas con QuitoMax® en el rendimiento y calidad de plántulas de tomate variedades ESEN y L-43. *Terra Latinoamericana*, 39, 1-6. <https://doi.org/10.28940/terra.v39i0.803>
- Hernández-Valencia, C. G., Román-Guerrero, A., Aguilar-Santamaría, Á., Cira, L., & Shirai, K. (2019a). Cross-linking chitosan into hydroxypropylmethylcellulose for the preparation of neem oil coating for postharvest storage of pitaya (*Stenocereus pruinosus*). *Molecules*, 24(2), 219. <https://doi.org/10.3390/molecules24020219>
- Hernández-Salido, L., & Salido-García, Y. (2019b). Influencia de la aplicación de Azofert inoculante a base *Rhizobium* en el cultivo del frijol común (*Phaseolus vulgaris* L.) var. Delicias 364 en finca Juan Sáez. Manatí. *Revista Caribeña de Ciencias Sociales*, 2019.
- Humber, R. A. (2008). Evolution of entomopathogenicity in fungi. *Journal of Invertebrate Pathology*, 98(3), 262-266. <https://doi.org/10.1016/j.jip.2008.02.017>
- Ibraheem, B. M. I., Hamed, S. M., Abd Elrhman, A. A., Farag, M. F., & Abdel-Raouf, N. (2017). Antimicrobial activities of some brown macroalgae against some soil borne plant pathogens and in vivo management of *Solanum melongena* root diseases. *Australian Journal of Basic and Applied Sciences*, 11(5), 157-68.
- INCA (Instituto Nacional de Ciencias Agrícolas). (2023). Productos, Cuba. Consultado el 27 de marzo, 2023, desde <https://www.inca.edu.cu/productos/>
- Ismail, E. G., Mohamed, W. W., Khattab, S., & Sherif, F. E. (2014). Effect of Manure and Bio-fertilizers on Growth, Yield, Silymarin content, and protein expression profile of *Silybum marianum*. *International Journal of Medicinal and Aromatic Plants*, 3(4), 430-438. <https://doi.org/10.15192/PSCP.AAB.2014.1.1.Article7>
- Jannet, H. B., H-skhir, F., Mighri, Z., Simmonds, M. S. J., & Blaney, W. M. (2001). Antifeedant activity of plant extracts and of new natural diglyceride compounds isolated from *Ajuga pseudoiva* leaves against *Spodoptera littoralis* larvae. *Industrial Crops and Products*, 14(3), 213-222. [https://doi.org/10.1016/S0926-6690\(01\)00086-3](https://doi.org/10.1016/S0926-6690(01)00086-3)
- Jaramillo, J. L., Montoya, E. C., Benavides, P., & Góngora, B. C. E. (2015). *Beauveria bassiana* y *Metarhizium anisopliae* para el control de broca del café en frutos del suelo. *Revista Colombiana de Entomología*, 41(1), 95-104.
- Katiyar, D., Hemantaranjan, A., Singh, B., & Bhanu, A. N. (2014). A future perspective in crop protection: Chitosan and its oligosaccharides. *Advances in Plants & Agriculture Research*, 1(1), 1-8.
- Kumar, R. S., Ayyadurai, N., Pandiaraja, P., Reddy, A. V., Venkateswarlu, Y., Prakash, O., & Sakthivel, N. (2005). Characterization of antifungal metabolite produced by a new strain *Pseudomonas aeruginosa* PUPa3 that exhibits broad-spectrum antifungal activity and biofertilizing traits. *Journal of Applied Microbiology*, 98(1), 145-154. <https://doi.org/10.1111/j.1365-2672.2004.02435.x>
- Lacey, L. A., Wraight, S. P., & Kirk, A. A. (2008). Entomopathogenic fungi for control of *Bemisia tabaci* biotype B: Foreign Exploration, Research and Implementation. In J. Gould, K. Hoelmer, J. Goolsby, (Eds.). *Classical Biological Control of Bemisia tabaci in the United States - A Review of Interagency Research and Implementation* (pp. 33-69). Dordrecht, Netherlands: Springer. [https://doi.org/10.1007/978-1-4020-6740-2\\_3](https://doi.org/10.1007/978-1-4020-6740-2_3)
- Lamz-Piedra, A., & González-Cepero, M. C. (2013). La salinidad como problema en la agricultura: la mejora vegetal una solución inmediata. *Cultivos Tropicales*, 34(4), 31-42.
- Lara, A. D., Costales, M. D., & Falcón, R. A. (2018). Los oligogalacturonidos en el crecimiento y desarrollo de las plantas. *Cultivos Tropicales*, 39(2), 27-134.
- Leon-Tacca, B., Ortiz-Calcina, N., Condori-Ticona, N., & Chura-Yupanqui, E. (2018). Cepas de *Trichoderma* con capacidad endofítica sobre el control del mildiu (*Peronospora variabilis* Gäum.) y mejora del rendimiento de quinua. *Revista de Investigaciones Altoandinas*, 20(1), 19-30. <https://dx.doi.org/10.18271/ria.2018.327>
- Lizárraga-Paulín, E. G., Torres-Pacheco, I., Moreno-Martínez, E., & Miranda-Castro, S. P. (2011). Chitosan application in maize (*Zea mays*) to counteract the effects of abiotic stress at seedling level. *African Journal of Biotechnology*, 10(34), 6439-6446.
- Logan, B., & Rabaey, K. (2012). Conversion of wastes into bioelectricity and chemicals by using microbial electrochemical technologies. *Science*, 337(6095), 686- 690.
- López-Llano, R. A., & Soto-Giraldo, A. (2016). Aislamiento de nematodos entomopatógenos nativos en cultivos de caña panelera y pruebas de patogenicidad sobre *Diatraea saccharalis* (Lepidoptera: Crambidae). Boletín Científico. Centro de Museos. *Museo de Historia Natural*, 20(2), 114-123. <https://doi.org/10.17151/bccm.2016.20.2.8>
- Malpartida-Zevallos, J., Narrea-Cango, M., & Dale-Larraburre, W. (2013). Patogenicidad de *Beauveria bassiana* (Bals) Vuill., sobre el gusano defoliador del maracuyá *Dione juno* (Cramer) (Lepidoptera: Nymphalidae) en laboratorio. *Ecología Aplicada*, 12(2), 75-81.
- Malusá, E., Sas-Paszt, L., & Ciesielska, J. (2012). Technologies for beneficial microorganisms inocula used as biofertilizers. *The Scientific World Journal*, 2012(1), 491206. <https://doi.org/10.1100/2012/491206>
- Mamani de Marchese, A., & Fillipone, M. P. (2018). Bioinsumos: componentes claves de una agricultura sostenible. *Revista Agronómica del Noroeste Argentino*, 38(1), 9-21.
- Martínez, A. (2012). Los plaguicidas botánicos y su importancia en la agricultura orgánica. *Revista Agricultura Orgánica*, 2(5), 26-30.
- Mascarin, G. M., & Jaronski, S. T. (2016). The production and uses of *Beauveria bassiana* as a microbial insecticide. *World Journal of Microbiology and Biotechnology*, 32, 1-26. <https://doi.org/10.1007/s11274-016-2131-3>
- Melgarejo, P., Romagosa, I., & Durán-Vila, N. (2014). Biotecnología agrícola. *Arbor*, 190(768), 152-152.
- Misra, N., Gupta, G., & Jha, P. N. (2012). Assessment of mineral phosphate-solubilizing properties and molecular characterization of zinc-tolerant bacteria. *Journal of Basic Microbiology*, 52(5), 549-558. <https://doi.org/10.1002/jobm.201100257>

- Morales-Guevara, D., Dell Amico-Rodríguez, J., Jerez-Mompié, E., Hernández, Y. D., & Martín-Martín, R. (2016). Efecto del QuitoMax® en el crecimiento y rendimiento del frijol (*Phaseolus vulgaris* L.). *Cultivos Tropicales*, 37(1), 142-147.
- Moreno, R. A., Carda, M. V., Reyes, C. J. L., Vásquez, A. J., & Cano, R. P. (2018). Rizobacterias promotoras del crecimiento vegetal: una alternativa de biofertilización para la agricultura sustentable. *Revista Colombiana de Biotecnología*, 20(1), 68-83. <https://doi.org/10.15446/rev.colomb.biote.v20n1.73707>
- Nava-Pérez, E., García-Gutiérrez, C., Camacho-Báez, J. R., & Vázquez-Montoya, E. L. (2012). Bioplaguicidas: una opción para el control biológico de plagas. *Ra Ximhai*, 8(3), 17-29.
- Ochoa-Villarreal, M., Vargas-Arispuro, I., Islas-Osuna, M. A., González-Aguilar, G., & Martínez-Téllez, M. Á. (2011). Pectin-derived oligosaccharides increase color and anthocyanin content in Flame Seedless grapes. *Journal of the Science of Food and Agriculture*, 91(10), 1928-1930. <https://doi.org/10.1002/jsfa.4412>
- Ortiz-Urquiza, A., & Keyhani N. O. (2013). Action on the surface: entomopathogenic fungi versus the insect cuticle. *Insects*, 4(3), 357-374.
- Páramo-Aguilera, L. A., Fonseca-Cruz, E. J., Delgado-Silva, H. D., Cabistán-Calderón, K. E., & Ríos-Guevara, C. K. (2018). La bioprospección en Nicaragua: avances en la búsqueda de aplicaciones agrícolas, industriales y ambientales. *Nexo Revista Científica*, 31(2), 89-103. <https://doi.org/10.5377/nexo.v31i2.6833>
- Plana-Llerena, R. R., González-Cañizares, P. J., & Soto-Carreño, F. (2016). Uso combinado de Ecomic®, Fitomas-e® y fertilizantes minerales en la producción de forraje para la alimentación animal a base de triticale (x. Triticosecale Wittmack), cv INCA TT-7. *Cultivos Tropicales*, 37(4), 76-83. <https://dx.doi.org/10.13140/RG.2.2.34452.30087>
- PLM® DeAQ/DIPO. (2023). Productos orgánicos/E-Microcyme. Consultado el 28 de marzo, 2023, desde <https://www.agroquimicos-organicosplm.com/emicrozyme/422/1/16029/152/9>.
- Ramos-Hernández, L., Arozarena-Daza, N. J., Lescaille-Acosta, J., García-Cisneros, F., Tamayo-Aguilar, Y., Castañeda-Hidalgo, E., ... & Rodríguez-Ortiz, G. (2013). Dosis de pectimorf® para enraizamiento de esquejes de guayaba var. Enana Roja Cubana. *Revista Mexicana de Ciencias Agrícolas*, 4(6), 1093-1105. <https://doi.org/10.29312/remexca.v0i6.1274>
- Ravikumar, S., Shanthi, S., Kalaiarasi, A., & Sumaya, M. (2014). The biofertilizer effect of halophilic phosphate solubilising bacteria on *Oryza sativa*. *Middle-East Journal of Scientific Research*, 19(10), 1406-1411 <https://doi.org/10.5829/idosi.mejrs.2014.19.10.11529>
- Reyes-Pérez, J. J., Rivero-Herrada, M., Solórzano-Cedeño, A. E., Carballo-Méndez, F. de J., Lucero-Vega, G., & Ruiz-Espinoza, F. H. (2021). Aplicación de ácidos húmicos, quitosano y hongos micorrízicos como influyen en el crecimiento y desarrollo de pimienta. *Terra Latinoamericana*, 39, 1-13. <https://doi.org/10.28940/terra.v39i0.833>
- Rivera, E. R. A., Fernández, M. F., Ruiz, M. L., González, C. P. J., Rodríguez, Y. Y., Ortega, P. E., ... & Lara F. D. (2020). Manejo, integración y beneficios del biofertilizante micorrízico EcoMic® en la producción agrícola. Mayabeque, Cuba: Instituto Nacional de Ciencias Agrícolas. ISBN: ISBN: 9789597258056
- Rodríguez-Pedroso, A. T., Ramírez-Arrebató, M. Á., Falcón-Rodríguez, A., Bautista-Baños, S., Ventura-Zapata, E., & Valle-Fernández, Y. (2017). Efecto del Quitomax® en el rendimiento y sus componentes del cultivar de arroz (*Oryza sativa* L.) var. INCA LP 5. *Cultivos Tropicales*, 38(4), 156-159.
- Rojas, R. L. (2013). Los basidiomicetos: una herramienta biotecnológica promisoría con impacto en la agricultura. *Fitosanidad*, 17(1), 49-55.
- Rumbos, C. I., & Athanassiou, C. G. (2017). The use of entomopathogenic nematodes in the control of stored-product insects. *Journal of Pest Science*, 90, 39-49.
- Salantur, A., Ozturk, A., & Akten, S. (2006). Growth and yield response of spring wheat (*Triticum aestivum* L.) to inoculation with rhizobacteria. *Plant Soil Environ*, 52(3), 111-118. <https://doi.org/10.17221/3354-PSE>
- Sánchez-Fernández, R. E., Sánchez-Ortiz, B. L., Sandoval-Espinosa, Y. K. M., Ulloa-Benítez, Á., Armendáriz-Guillén, B., García-Méndez, M. C., & Macías-Rubalcava, M. L. (2013). Hongos endófitos: fuente potencial de metabolitos secundarios bioactivos con utilidad en agricultura y medicina. *TIP Revista Especializada en Ciencias Químico-Biológicas*, 16(2), 132-146.
- Sánchez, J. J., Valle, D. J., Pérez, T. E., Neira, de P. M., & Calderón, A. C. (2019). Biological control of *Spodoptera frugiperda* in *Zea mays* culture: Use of entomopathogenic nematodes. *Scientia Agropecuaria*, 10(4), 551-557. <https://dx.doi.org/10.17268/sci.agropecu.2019.04.12>
- Seipasa (2023). Bioestimulantes agrícolas. Consultado el 28 de marzo, 2023, desde [https://www.seipasa.com/es\\_ES/bioestimulantes/](https://www.seipasa.com/es_ES/bioestimulantes/)
- Sharma, H. S., Fleming, C., Selby, C., Rao, J. R., & Martin, T. (2014). Plant biostimulants: a review on the processing of macroalgae and use of extracts for crop management to reduce abiotic and biotic stresses. *Journal of Applied Phycology*, 26, 465-490.
- Syngenta (2023). Productos y soluciones. Consultado el 30 de marzo, 2023, desde <https://www.syngenta.com.mx/products/search/crop-protection>
- Tamayo, A. Y., Riera, N. N., Terry, A. E., Juárez, L. P., & Rodríguez, M. Y. (2019). Respuesta de *Vigna unguiculata* (L) Walp a la aplicación de bioproductos en condiciones de huertos intensivos. *Acta Agronómica*, 68(1), 41-46. <https://doi.org/10.15446/acag.v68n1.72797>
- Terry-Alfonso, E., Ruiz-Padrón, J., & Carrillo-Sosa, Y. (2018). Efecto de diferentes manejos nutricionales sobre el rendimiento y calidad de frutos de tomate. *Agronomía Mesoamericana*, 29(2), 389-401. <https://doi.org/10.15517/ma.v29i2.28889>
- Tirado-Gallego, P. A., Lopera-Álvarez, A., & Ríos-Osorio, L. A. (2016). Estrategias de control de *Moniliophthora roreri* y *Moniliophthora perniciosa* en *Theobroma cacao* L.: revisión sistemática. *Ciencia y Tecnología Agropecuaria*, 17(3), 417-430. [https://doi.org/10.21930/rcta.vol17\\_num3\\_art:517](https://doi.org/10.21930/rcta.vol17_num3_art:517)
- Velasco-Jiménez, A., Castellanos-Hernández, O., Acevedo-Hernández, G., Aarland R. C., & Rodríguez-Sahagún, A. (2020). Bacterias rizosféricas con beneficios potenciales en la agricultura. *Terra Latinoamericana*, 38(2), 333-345. <https://doi.org/10.28940/terra.v38i2.470>
- Viera-Arroyo, W. F., Tello-Torres, C. M., Martínez-Salinas, A. A., Navia-Santillán, D. F., Medina-Rivera, L. A., Delgado-Párraga, A. G., ... & Jackson, T. (2020). Control Biológico: Una herramienta para una agricultura sustentable, un punto de vista de sus beneficios en Ecuador. *Journal of the Selva Andina Biosphere*, 8(2), 128-149.
- Villarreal-Delgado, M. F., Villa-Rodríguez, E. D., Cira-Chávez, L. A., Estrada-Alvarado, M. I., Parra-Cota, F. I., & Santos-Villalobos, S. D. L. (2018). The genus *Bacillus* as a biological control agent and its implications in the agricultural biosecurity. *Revista Mexicana de Fitopatología*, 36(1), 95-130. <https://doi.org/10.18781/r.mex.fit.1706-5>
- Woo, S. L., Ruocco, M., Vinale, F., Nigro, M., Marra, R., Lombardi, N., ... & Lorito, M. (2014). *Trichoderma*-based products and their widespread use in agriculture. *The Open Mycology Journal*, 8(1). <http://dx.doi.org/10.2174/1874437001408010071>
- Wraight, S. P., & Ramos, M. E. (2002). Application parameters affecting field efficacy of *Beauveria bassiana* foliar treatments against Colorado potato beetle *Leptinotarsa decemlineata*. *Biological Control*, 23(2), 164-178. <https://doi.org/10.1006/bcon.2001.1004>